

FOIANI MARTHA¹, Jackson-Morgan T.², Cicognola C.³, Ermann N.⁴, Heslegrave A.¹, Ye, K.⁵, Kornhuber, J.⁴, Lewczuk P.^{4,6}, Zetterberg H.^{1,3}, Hoglünd K³, Rohrer J.D.⁷, Lashley T.²

Prof Henrik Zetterberg

¹Dementia Research Institute, University College London, London, UK; ²Queen Square Brain Bank (QSBB), University College London, London, UK; ³University of Gothenburg, Gothenburg, Sweden; ⁴Department of Psychiatry and Psychotherapy, Universitätsklinikum Erlangen, and Friedrich-Alexander-Universität Erlangen-Nürnberg, Erlangen, Germany; ⁵Pathology & Laboratory Medicine, Experimental Pathology, Emory University School of Medicine, Atlanta, Georgia, USA; ⁶Department of Neurodegeneration Diagnostics, Medical University of Białystok, Białystok, Poland; ⁷Dementia Research Centre, University College London, London, UK

Background

Around 40% of patients with frontotemporal dementia (FTD) have tau-positive inclusions at post-mortem with a variety of different pathologies found, named primary tauopathies. Unique conformations of tau are hypothesized to underlie the distinct morphological and cellular distribution of pathological tau aggregates. In this study we investigated the potential of a set of antibodies (Fig 1) targeting the whole length of the tau protein to detect the different conformational changes in the primary tauopathies (Fig 2). The aim of the work was to identify distinct conformational changes that could be detected with specific antibodies and facilitate a post mortem diagnosis for the pathology.

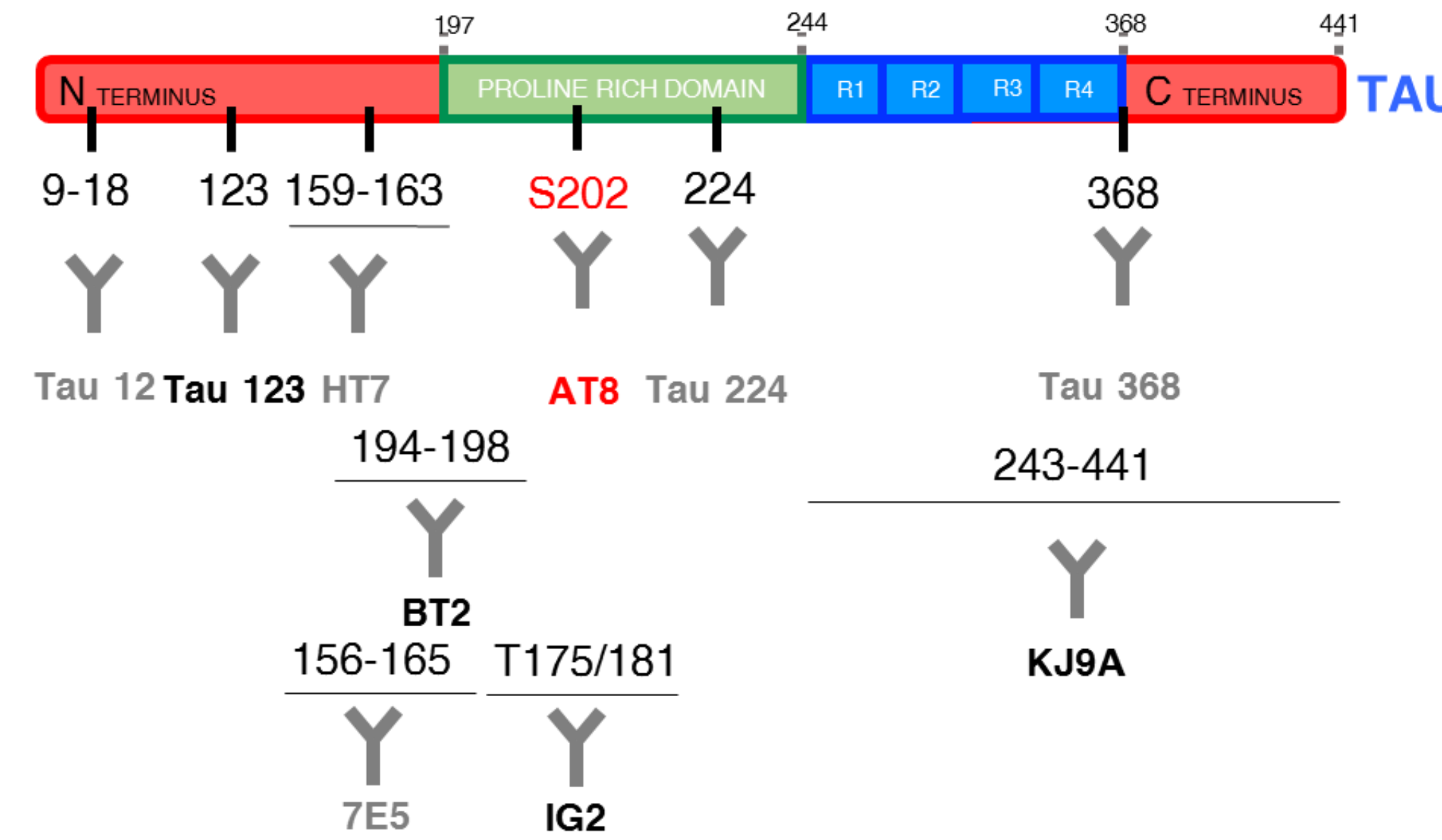


Figure 1: Schematic of tau 441 protein with the approximate location of various linear epitope antibodies.

Methods

Brain tissue from twenty two cases of primary tauopathy from the Queen Square Brain Bank (QSBB), London, UK were assessed (Table 1): five each with Pick's disease (PiD), progressive supranuclear palsy (PSP), corticobasal degeneration (CBD), and two with globular glial tauopathy (GGT) as well as five cases of Alzheimer's disease for comparison. Eight μ m sections from paraffin wax blocks of brain tissue from the frontal cortex, temporal cortex and hippocampus of each case were selected. Ten different antibodies were used: four targeting the N-terminal (tau 123, tau 12, HT7 and 7E5), three targeting the proline-rich domain (BT2, tau 224 and IG2) and two targeting the C-terminal (tau 368 and KJ9A) (Fig 1).

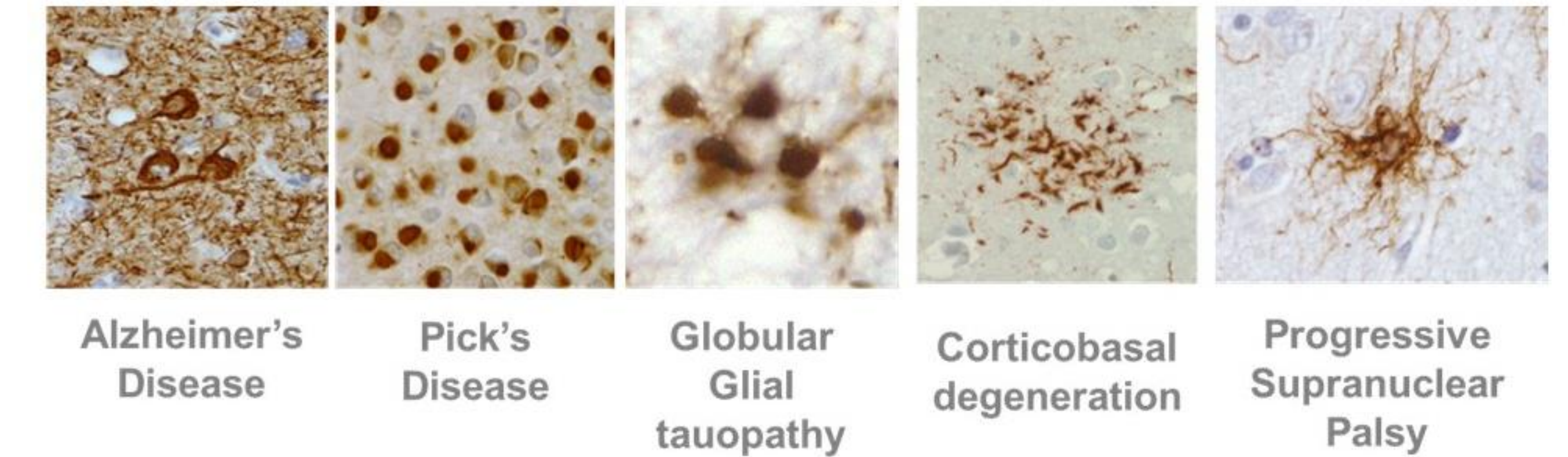


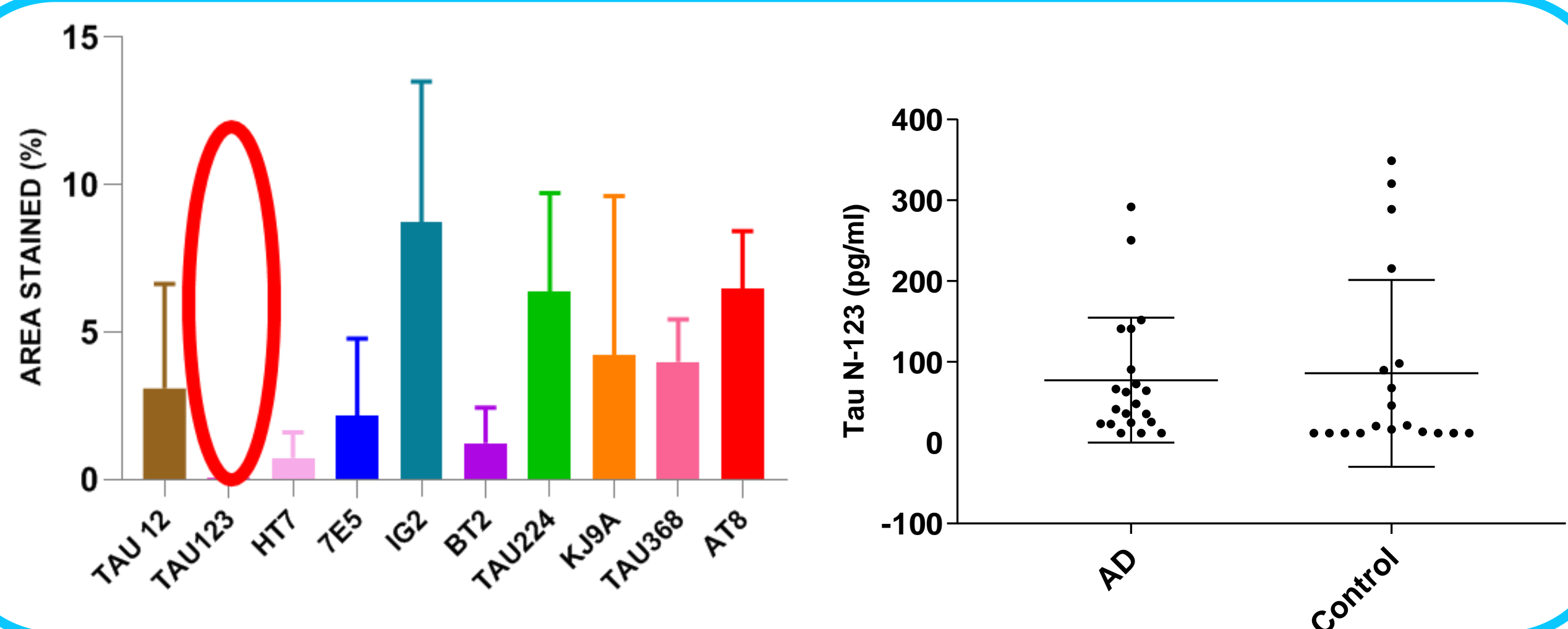
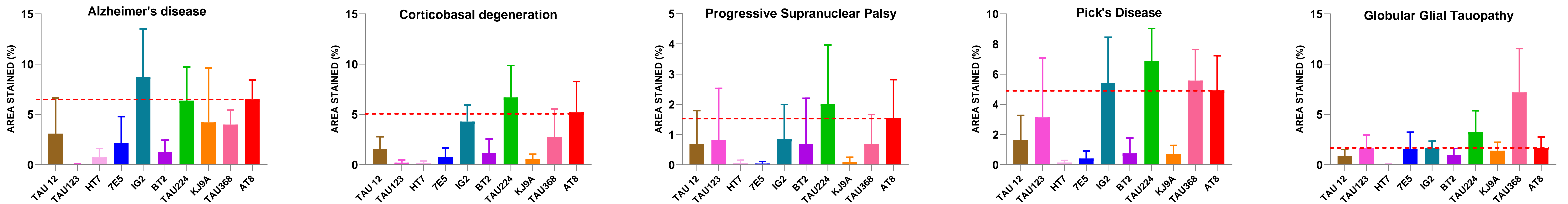
Figure 2: Representative images of pathological tau inclusions in IHC stained post mortem tissue from different tauopathies.

Pathological diagnosis	Number	Age	Gender (M%)	Disease Duration
PiD	5	60	60	7
PSP	5	75	40	9
CBD	5	57	60	7
GGT	2	78	50	5
AD	5	72	40	16

Table 1: Table containing the demographical data of pathological cases included in the study

Results

Results show that individual antibodies selectively bind to specific tauopathies whilst staining negative to others, confirming a distinct fibril organisation for each tauopathy. Comparing all results to the gold standard for staining of tau pathological inclusions (using AT8 antibody), N-terminal antibodies mostly stained PiD inclusions and PSP astrocytic plaques. Antibodies targeting the proline rich domain of tau, positively stained the pathological inclusions present in AD, PSP, PiD and GGT although tau 224 stained pathological tau in all pathologies. Finally, C-terminal antibodies positively bound to astrocytic inclusions present in CBD and PSP.



The results obtained were confirmed with an immunoassay approach measuring the Tau 9-123 and Tau 9-224 fragments (ELISA and Simoa HD-1 Platform homebrew assay respectively). Tau 123 did not stain inclusions present in Alzheimer's disease and the same protein could not be detected in CSF of patients compared to controls. Tau 224 stained primary tauopathies inclusions as strongly as AT8 and CSF measurement of tau 2224 showed a significant difference between FTD and controls.

Conclusions

Here, we report the potential of several tau antibodies to selectively bind to specific forms of tauopathy.

